

# The User's Manual—MDCK SFM01

## **Basic Information Introduction**

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MDCK SFM01 is serum-free media, without serum, protein, any animal-derived components and ect, formulated with high quality raw materials, high quality and stability, and no serum is required for the culture process, it suitable for high-density cell suspension expansion of MDCK cell, supporting MDCK rapid expansion, high-density culture and influenza virus vaccine production under a completely serum-free protein-free system.



### **Application Scope**

MDCK SFM01 medium is suitable for suspension domestication and large-scale serum-free suspension culture of MDCK cells. It can be used in the development and production process of influenza virus vaccine and other biological products, and can support the development to large-scale production, but cannot be directly used in the human body or used as a medication.

### Shipping, Storage and Validity Period

Product	Catalog No.	Storage	Shipping	Validity Period
MDCK SFM01	LQ17, Liquid	2°C ~ 8°C, Protect from light	2°C ~ 8°C, Protect from light	12 months
MDCK SFM01	DP17, Powder	2°C ~ 8°C, Protect from light	2°C ~ 8°C, Protect from light	24 months

### **Protocol for Hydration of Powder Medium**

- 1. Fill the mixing container with purified water  $(28 \sim 32^{\circ}C)$  at 100% of the final volume.
- 2. Slowly add 27.21 g/L of powder medium with gentle stirring. Mix for  $18 \sim 22$  minutes.
- 3. Add 5 mol/L NaOH solution to the above solution at  $0.25 \pm 0.01$  g/L. Mix for  $18 \sim 22$  minutes.
- 4. Slowly add 2.00  $\pm$  0.05 g/L NaHCO3 to the solution. Mix for 18  $\sim$  22 minutes.
- 5. Filter immediately the media with a 0.22  $\mu m$  membrane filter.



## Version 2, 2023

Product Index	MDCK SFM01 (LQ19), Liquid	MDCK SFM01 (DP19), Powder	
Appearance	Yellow, clear liquid	Light yellow or similar color powder	
pH	$7.0 \sim 7.4$	7.0 ~ 7.4 (pre-filter)	
Osmolality (mOsmol/kg)	290 ~ 355	300 ~ 350	
Solubility		Dissolve well according to the protocol for hydration of powder medium	
Endotoxin (EU/mL)	< 50	< 50	
Sterility	Negative		
Bioburden		Aerobic bacteria: < 200 CFU/g Molds and yeasts: < 200 CFU/g	

## **Quality Index of Powder and Liquid Media**

# **Reference Cell Culture Protocol**

# **Culture Conditions**

Parameter		Value	
Culture volume	50 mL TPP Tube	10 ~ 30 mL	
	125 mL Shake flask	15 ~ 40 mL	
	250 mL Shake flask	40 ~ 80 mL	
	500 mL Shake flask	100 ~ 200 mL	
	1000 mL Shake flask	200 ~ 300 mL	
Shaking speed	TPP Tube	50mm amplitude: 200 rpm	
	Shake flask	25mm amplitude: 150 rpm	
	Shake flask	50mm amplitude: 90 ~ 120 rpm	
Culture environment	Seeding density	1.0×10 <sup>6</sup> cells/mL	
	Incubation temperature	37°C	
	Incubation CO <sub>2</sub> concentration	5%	
	Incubation relative humidity	> 80% RH	



### **Cell Thawing**

1. Pre-warm the medium (MDCK SFM01) in 37°C water bath.

2. Spray the outside of the medium bottle with 75% alcohol and place the bottle into the bio-safety cabinet.

3. Thaw one vial at a time in 37°C water bath. Gently agitate the vial within 1 minutes until the ice in the vial melting.

4. Pipet the contents from the vial gently into a centrifuge tube containing 30 mL of pre-warmed medium (MDCK SFM01).

5. Centrifuge 170 g ~ 190 g for 5 minutes. Discard the supernatant and re-suspend cells in 10 mL fresh pre-warmed medium (MDCK SFM01), then adjust the cell density to  $1 \times 10^{6}$  cells/mL.

6. Sample 0.5 mL of cell suspension and analyze the viable cell density ( $\times 10^6$  cells/mL) and viability (%) of the sample using cell counter.

7. If the cell viability > 90%, incubate cells in the specified condition (refer to "culture conditions" table).

### **Cell Passage**

1. Pre-warm the medium (MDCK SFM01) in  $37^{\circ}$ C water bath for  $20 \sim 30$  min.

2. Cells with viable cell density  $\ge 4 \times 10^6$  cells/mL, cell viability  $\ge 90\%$ , and in the middle of logarithmic growth phase were selected for passage.

3. According to the seed cell density of  $1.0 \times 10^6$  cells/mL, calculate the amount of total number of seed cells.

4. Seed cells at  $1.0 \times 10^6$  cells/mL in the flask and add a certain volume of pre-warmed fresh medium.

5. Incubate cells in the specified environment condition (refer to "culture conditions" table).

6. Passage cells with fresh MDCK SFM01 medium according to the above steps once every  $48 \pm 3$  hours.

7. If the viable cell density is less than  $4.0 \times 10^6$  cells/mL or the viability is lower than 90% before passaging, the cells need to be centrifuged at 170 g ~ 190 g for 5 minutes. Carefully remove the spent media, then resuspend cells with preheated MDCK SFM01 medium, passage cells after sampling and counting.

## Adaptation

#### **Direct Adaptation**

1. For cells can direct adapt, transfer cells suspension cultures into MDCK SFM01 directly, and the seed cell density refer to the cell passage procedure.

2. Cell passage until cell expression steadily.

3. When VCD reaches  $4 \times 10^6$  cells/mL and > 90% viability ( $48 \pm 3$  hours). At this point, the cells had been successful



adapted.

#### **Sequential Adaptation**

For cells growing in  $5 \sim 10\%$  serum or SFM media. Sequential adaptation should be performed.

1. Seed cells at  $1.0 \times 10^6$  cells/mL in original cell culture media.

2. Sample and cell count every day until the VCD reaches  $4 \times 10^6$  cells/mL.

3. Seed cell density at  $1.0 \times 10^6$  cells/mL, subculture cells into stepwise increasing ratios of complete MDCK SFM01 medium to original medium with each subsequent passage (25:75, 50:50, 75:25, 90:10, 100:0).

4. When VCD reaches  $4 \times 10^6$  cells/mL and  $\geq 90\%$  viability ( $48 \pm 3$  hours). At this point, the cells had fully adapted to MDCK SFM01 media.

### **Cell Cryopreservation**

1. Prepare cells, harvesting in mid-log phase of growth with viability > 90%.

2. Sample and cell counting, calculate the required volume of cell freeze solution to give a final density of  $1 \times 10^7$  cells/mL.

3. Prepare the cell freeze solution: 93% MDCK SFM01 + 7% DMSO, store at 4°C.

4. Centrifuge  $170 \text{ g} \sim 190 \text{ g}$  for 5 minutes, discard the supernatant and re-suspend cells with the cell freeze solution.

5. Immediately dispense aliquots of cells suspension into cryovials according to the specific needs of the project.

6. Achieve cryopreservation in an automated or manual controlled rate freezing apparatus following standard procedures (1°C decrease per minute).

7. Transfer to liquid nitrogen tank for storage.